

Synthesis of 26-Iodoponasterone, a New and very Active Ecdysteroid

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26-Iodoponasterone A, prepared from inokosterone in four steps, is 160 times as active as the parent compound in an ecdysone assay on *Drosophila Kc* cells, and is one of the most active ecdysones known; the ¹²⁵I is being used for the detection of ecdysone receptors.

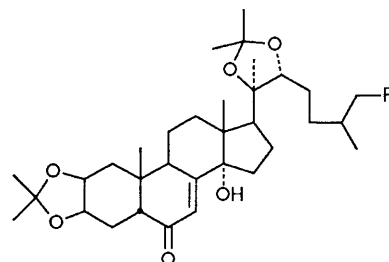
20-Hydroxyecdysone, the moulting hormone of arthropods, like other steroids, is a regulator of gene expression, and the molecular biology of its actions is being studied in numerous laboratories.^{1,2,3} Work on ecdysone receptors requires the use of hormone analogues more active than the natural hormone 20-hydroxyecdysone,^{2,4,5} and the development of potent ecdysteroids suitable for radiolabelling has been critical to progress in this field. Here, we report the synthesis of a new ecdysone analogue, 26-iodoponasterone, starting from the phytoecdysone inokosterone **1**. 26-Iodoponasterone proves to be a very active ecdysone suitable for use in receptor studies and for radiolabelling.⁶

Inokosterone was isolated from the Chinese herb *Achyranthus fauriei* following published procedures.⁷ Treatment of inokosterone (**1**, 53 mg) with dry acetone (3 ml) and anhydrous *p*-TsOH (5 mg) yielded 45 mg of the diacetonide (**2**, 87% yield) after work-up and purification through a silica gel column (5 g), elution with 5% EtOH-CHCl₃. The product **2** shows the following NMR signals: δ (CDCl₃, 250 MHz) 5.80 (d, *J* 2.1 Hz, 7-H), 4.24 (m, 3-H), 4.20 (m, 2-H), 3.47 (d, *J* 5.2 Hz, 26-H₂), 2.78 (dt, *J* 8.1, 8.1 and 2.1 Hz, 9-H), 2.33 (dd, *J* 12.5 and 3.7 Hz, 5-H), 1.47 and 1.30 (2,3-acetonide-Me), 1.38 and 1.30 (20,22-acetonide-Me), 1.12 (20-Me), 0.96 (s, 10-Me), 0.93 and 0.91 (d, *J* 6.5 Hz, 25-Me) and 0.77 (13-Me). The 26-hydroxy of the diacetonide **2** (25 mg) was mesylated by dropwise treatment with mesyl chloride (0.1 ml) in dry pyridine (2 ml) at 0 °C for 0.5 h and at ambient temperature for 1 h. After addition of pyr-H₂O (2:1, 1 ml), the solution was partitioned between H₂O and Et₂O. The residue of the organic layer (24 mg) was purified by silica gel (5 g) chromatography, elution with 1 and 2.5% EtOH-CHCl₃, to give 19 mg of mesylate **3**; Me of mesyl group at δ 3.02, and 26-H₂ at δ 4.10.

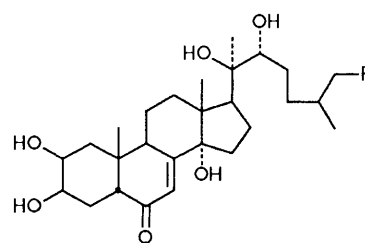
The iodo derivative **4** was prepared by treating 5 mg mesylate **3** with 28 mg Bu₄NI in 2 ml toluene at 75 °C under argon for 19 h.⁸ The cooled reaction mixture was then partitioned three times⁸ between 30 ml H₂O and 30 ml ether, and the residue of the ether layer (7 mg) was purified with silica gel column (3 g), elution with 1% EtOH-CHCl₃ to give 6 mg of the iodo product; NMR δ (CDCl₃): 5.83 (d, *J* 2.0 Hz), 4.27 (m, 3-H), 4.23 (m, 2-H), 3.25 (m, 26-H₂), 2.80 (ddd, *J* 8.5, 6.4 and 2.0 Hz, 9-H), 2.36 (dd, *J*, 12.5 and 4.6 Hz, 5-H), 1.50 and 1.33 (s, 2,3-acetonide-Me), 1.41 and 1.33 (s, 20,22-acetonide-Me), 1.13 (s, 20-Me), 1.02 (d, *J* 6.3 Hz, 25-Me), 0.99 (s, 10-Me), and 0.79 (s, 13-Me). The 20,22-acetonide, which is relatively stable toward HCl-THF or HOAc-THF-H₂O pairs, can be removed readily treating with 10% HClO₄-MeOH (1:1). 26-Mesylinkosterone **5** or 26-iodoponasterone A **6** was obtained in 70% yield by overnight treatment of mesyldiacetonide **3** or iododiacetonide **4** with 10% HClO₄-MeOH (1:1), work-up (neutralization with NaHCO₃ then extraction with EtOAc) and purification (silica gel, eluent: 10, 15 and 20% EtOH-CHCl₃).

Mesylate **5** shows the following NMR signals: δ (CD₃OD) 5.80 (d, *J* 2.2 Hz, 7-H), 4.09 (m, 26-H₂), 3.94 (m, 3-H), 3.83 (dt, *J* 11.4, 3.9 and 3.9 Hz, 2-H), ~3.40 (overlapping with MeOH, 22-H), 3.13 (m, 9-H), 3.06 (s, mesyl-Me), 2.36 (dd, *J* 12.5 and 4.6 Hz, 5-H), 1.18 (s, 20-Me), 1.10 and 1.08 (d, *J* 6.6 Hz, 25-Me), 0.96 (s, 10-Me) and 0.88 (s, 13-Me). Spectral data

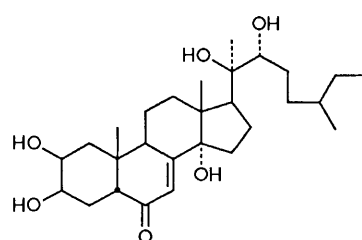
of iodoponasterone **6** are as follows: desorption chemical ionization (DCI) (NH₃ reactant gas) C₂₇H₄₃O₆I, *m/z* 590 (M⁺), λ_{max} (MeOH) 244 nm (ε 12 000); δ (CD₃OD) 5.80 (d, *J* 2.2 Hz, 7-H), 3.94 (m, 3-H), 3.38 (m, 2-H), 3.40 (overlapping with MeOH, 22-H), 3.22 (m, 26-H₂), 3.14 (m, 9-H), 2.36 (dd, *J* 12.5 and 4.6 Hz, 5-H), 1.14 (s, 20-Me), 1.00 and 0.98 (d, *J* 5.6 Hz, 25-Me), 0.96 (s, 10-Me) and 0.88 (s, 13-Me). As summarized in **6a**, the DCI-MS of **6** showed peaks at *m/z* 462 (37%), 444 (40%) and 427 (37%) resulting from the loss of HI, HI+H₂O and H+H₂O+OH, respectively. The chemical shifts of 26-H₂, δ 3.47 (in **2**, CH₂OH), 4.10 (in **3** and **5**, CH₂OMs), and 3.22 (in **4** and **6**, CH₂I) can be accounted for by the electronegativity differences of substituents on C-26; the mesylate NMR signals at δ 3.02 also agree with **3** and **5**.



2 R = OH
3 R = OMs
4 R = I



1 R = OH
5 R = OMs
6 R = I
7 R = H



6a

DCI-MS (NH₃): *m/z*

590 (M⁺, 37%)
572 (M⁺ -H₂O, 95%)
554 (M⁺ -2H₂O, 100%)
536 (M⁺ -3H₂O, 91%)
462 (M⁺ -HI, 37%)
444 (M⁺ -HI-H₂O, 40%)
427 (M⁺ -HI-H₂O-OH, 37%)

The HPLC retention times for **5** and **6** are 3.42 and 10 min, respectively, under the following conditions: 5 μ m analytical Ultrasphere ODS column, MeOH-H₂O (60:40) as eluent, 1 ml/min (flow rate) and detection at 242 nm.

We have examined the structure-activity relationships among inkosterone **1**, mesylate **5**, iodoponasterone A **6**, ponasterone A **7**, and 20-hydroxyecdysone (20-HE), using the standard *Drosophila* Kc cell assay, in which previous results have correlated well with known receptor affinities.^{6,9} Relative to 20-HE, the activities were **1** 0.1, **5** 0.1, **6** 16, and **7** 8. Thus, in the Kc cell bioassay, 26-iodoponasterone **6** is more potent than 20-HE by at least an order of magnitude, and is in fact more active than any previously described ecdysone, except the synthetic analogue 14-desoxymuristerone A.¹⁰ Procedures for the preparation of [¹²⁵I]26-iodoponasterone A of high specific activity from the mesylate **5** have been described previously.⁴

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References

- 1 L. Cherbas, H. Benes, M. Bourouis, K. Burtis, A. Chao, P. Cherbas, M. Crosby, M. Garfinkel, G. Guild, D. Hogness, J. Jami, C. W. Jones, M. Koehler, J.-A. Lepesant, C. Martin, F. Maschat, P. Mathers, E. Meyerwitz, R. Moss, R. Pictet, J. Rebers, G. Richards, J. Roux, R. Schulz, W. Segraves, C. Thummel and K. Vijayaraghavan, *Insect Biochem.*, 1986, **161**, 241.
- 2 H. J. Bidmon and T. J. Sliter, *Invertebr., Reprod. and Devel.*, 1990, **18**, 13.
- 3 W. A. Segraves and G. Richards, *Invertebr. Reprod. and Devel.*, 1990, **18**, 67.
- 4 P. Maroy, R. Dennis, C. Beckers, B. A. Sage and J. D. O'Connor, *Proc. Natl. Acad. Sci. USA*, 1978, **75**, 6035.
- 5 M. A. Yund, D. W. King and J. W. Fristrom, *Proc. Natl. Acad. Sci. USA*, 1978, **75**, 6039.
- 6 P. Cherbas, L. Cherbas, S.-L. Lee and K. Nakanishi, *Proc. Natl. Acad. Sci. USA*, 1988, **85**, 2096.
- 7 T. Takemoto, S. Ogawa and N. Nishimoto, *Yakugaku Zasshi*, 1967, **87**, 1463, 1469.
- 8 C. Ducrocq, A. Righini-Tapie, R. Azerad, J. F. Green, P. A. Freidman, J.-P. Beaucourt and B. Rousseau, *J. Chem. Soc., Perkin Trans. 1*, 1986, 1323.
- 9 L. Cherbas, C. D. Yonge, P. Cherbas and C. M. Williams, *Wilhelm Roux's Arch., Dev. Biol.*, 1980, **1**, 189.
- 10 P. Cherbas, D. A. Trainor, R. J. Stonard and K. Nakanishi, *J. Chem. Soc., Chem. Commun.*, 1982, 1307.